

Labeling the Stroma of a Patient-Derived Orthotopic Xenograft (PDOX) Mouse Model of Undifferentiated Pleomorphic Soft-Tissue Sarcoma With Red Fluorescent Protein for Rapid Non-Invasive Imaging for Drug Screening

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ABSTRACT

Our laboratory pioneered patient-derived orthotopic xenograft (PDOX) mouse models using surgical orthotopic implantation (SOI). PDOX models are patient-like, in contrast to the ectopic subcutaneous-transplant cancer models. In the present study, we demonstrate that an undifferentiated pleomorphic soft-tissue sarcoma (UPS-STS) PDOX model acquired bright RFP-expressing stroma through one passage in red fluorescent protein (RFP) transgenic mice, which upon passage to non-colored nude mice was non-invasively imageable. A PDOX nude mouse model of UPS-STS was established in the biceps femoris of nude mice. After the tumors grew to a diameter of 10 mm, the tumors were subsequently passaged to RFP transgenic mice, and after tumor growth were then passaged to non-transgenic nude mice. Tumors were divided into small fragments and transplanted in the biceps femoris at each passage. The OV100 Small Animal Fluorescence Imaging System and FV1000 laser scanning confocal microscope were used to image RFP fluorescence in the UPS-STS PDOX models. UPS-STS PDOX tumors, previously grown in RFP transgenic nude mice for only one passage, had very bright fluorescence and after passage to non-transgenic nude mice maintained the bright fluorescence and were non-invasively imageable. FV1000 confocal imaging revealed diffusely distributed bright RFP stromal cells in the PDOX tumor, both in RFP transgenic mice and after passage to non-transgenic mice. These results demonstrate a powerful method to make the PDOX UPS-STS model brightly fluorescent for non-invasive imaging, as well as for confocal microscopy of individual stromal cells associated with the tumor. The RFP-labeled UPS PDOX has the potential to rapidly screen for novel effective agents for individual patients, including stroma-targeting drugs, whereby the stromal cells are a visual target. *J. Cell. Biochem.* 118: 361–365, 2017. © 2016 Wiley Periodicals, Inc.

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Orthotopic implantation of intact tumor tissue in appropriate mouse models can result in metastasis resembling the clinical pattern, unlike subcutaneous transplantation [Hoffman, 2015]. We have established patient-derived orthotopic xenograft (PDOX) nude-mouse models of the following patient cancers: colon [Fu et al., 1991]; pancreas [Fu et al., 1992; Hiroshima et al., 2014a,b,c,d, 2015a, b; Yano et al., 2015]; lung [Wang et al., 1992]; ovarian [Fu and Hoffman, 1993]; breast [Fu et al., 1993]; stomach [Furukawa et al., 1993]; mesothelioma [Astoul et al., 1996]; soft tissue sarcoma [Hiroshima et al., 2015c,d; Murakami et al., 2016a]; follicular dendritic-cell sarcoma [Kiyuna et al., 2016]; and Ewing's sarcoma [Murakami et al., 2016b].

We previously developed an imageable PDOX model for pancreatic cancer [Suetsugu et al., 2012a]. Pancreatic cancer PDOX models were serially passaged to transgenic green fluorescent protein (GFP), red fluorescent protein (RFP), and cyan fluorescent protein (CFP), respectively. The PDOX acquired and maintained fluorescent stroma for each mouse. The PDOX, which acquired GFP-expressing stroma, subsequently metastasized to the liver and formed peritoneal metastases which both maintained the stroma from the primary tumor [Suetsugu et al., 2012b]. PDOX that acquired GFP and RFP stroma, were then orthotopically passaged to non-transgenic nude mice which enabled noninvasive longitudinal imaging as the tumor progressed in non-transgenic nude mice [Suetsugu et al., 2012c].

Previously, an undifferentiated pleiomorphic soft-tissue sarcoma (UPS) was established by our laboratory in the biceps muscle of nude mice and was eradicated with tumor-targeting *Salmonella typhimurium* A1-R followed by doxorubicin (DOX) [Murakami et al., 2016a].

The present report demonstrates a non-invasive imageable UPS PDOX model, established with brightly labeled RFP-expressing sarcoma through only a single passage through an RFP transgenic nude mouse.

MATERIALS AND METHODS

MICE

Athymic *nu/nu* nude mice and transgenic RFP expressing athymic *nu/nu* mice (AntiCancer Inc., San Diego, CA), 4–6 weeks old, were used in this study [Yang et al., 2009]. All surgical procedures and imaging were performed with an AntiCancer Institutional Animal Care and Use Committee (IACUC)-protocol specifically approved for this study, and in accordance with the principles and procedures outlined in the National Institute of Health Guide for the Care and Use of Animals under Assurance Number A3873-1. In order to minimize any suffering of the animals, the use of anesthesia and analgesics were used for all surgical experiments. Animals were anesthetized by subcutaneous injection of a 0.02 ml solution of 20 mg/kg ketamine, 15.2 mg/kg xylazine, and 0.48 mg/kg acepromazine maleate. The response of animals during surgery was monitored to ensure adequate depth of anesthesia. The animals were observed on a daily basis humanely sacrificed by CO₂ inhalation, if they met the following humane endpoint criteria: severe tumor burden (more than 20 mm in diameter), prostration, significant body

weight loss, difficulty breathing, rotational motion, and body temperature drop. Animals were housed in a barrier facility on a high efficiency particulate arrestance (HEPA)-filtered rack under standard conditions of 12-h light/dark cycles. The animals were fed an autoclaved laboratory rodent diet.

PATIENT-DERIVED TUMOR

The patient was diagnosed with UPS of the left thigh and had the tumor resected. The disease recurred locally a few months later and the patient was treated with radiotherapy, and subsequent re-resection by F.C.E., Division of Surgical Oncology, University of California, Los Angeles (UCLA). Written informed consent was obtained from the patient as part of a UCLA Institutional Review Board-approved protocol.

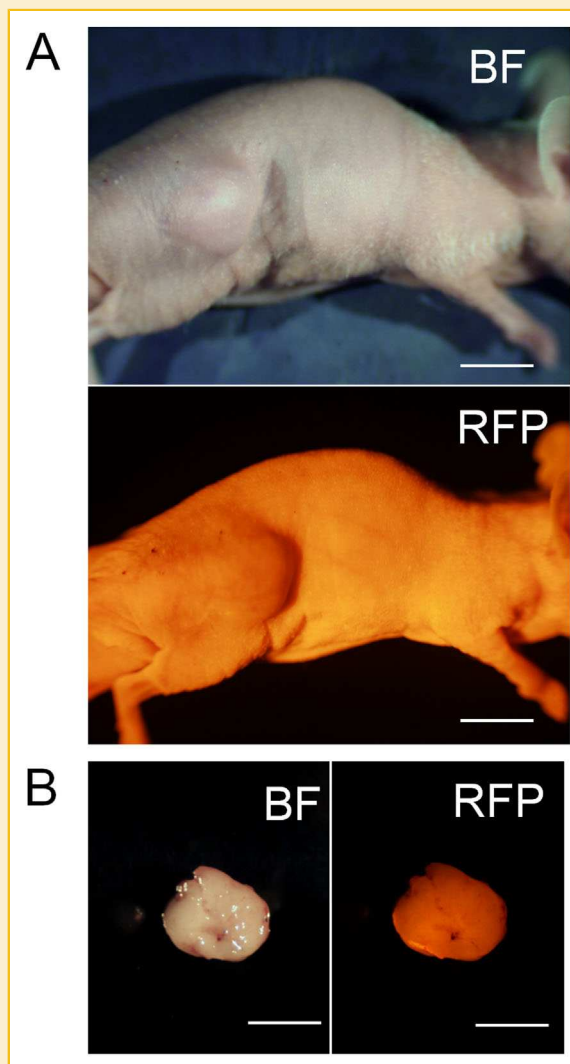


Fig. 1. Establishment of a fluorescent undifferentiated pleiomorphic soft-tissue sarcoma (UPS) PDOX model. (A) A growing PDOX tumor in the right biceps femoris of an RFP-expressing nude mouse. (B) Images of the resected tumor derived from the RFP-expressing nude mouse. Scale bars: 10 mm (A and B). RFP, red fluorescent protein; BF, brightfield.

ESTABLISHMENT OF A FLUORESCENT PDOX MODEL OF UPS BY SURGICAL ORTHOTOPIC IMPLANTATION (SOI)

A fresh sample of UPS was obtained immediately after patient surgery. The tumor was transported immediately to the laboratory at AntiCancer, Inc. on wet ice. The tumor was cut into fragments and implanted subcutaneously in nude mice. Two weeks later, the subcutaneously implanted tumors grew to more than 10 mm in diameter. The subcutaneously grown tumors were then harvested and cut into small fragments. A 5-mm skin incision was made on the high thigh into the biceps femoris, under ketamine anesthesia, which was split, and a single tumor fragment was implanted orthotopically in this space to establish PDOX model in RFP transgenic nude mice. The wound was closed with a 6-0 nylon suture (Ethilon, Ethicon, Inc., NJ). When the PDOX tumors in the RFP transgenic mice grew to more than 10 mm in diameter, the tumor was harvested and passaged to non-transgenic nude mice with the same method described above.

IMAGING OF THE FLUORESCENT UPS PDOX MODEL

Fluorescence imaging of the macroscopic tumor was performed with the OV100 Small Animal Imaging System (Olympus, Tokyo, Japan) [Yamauchi et al., 2006]. When PDOX tumors reached 10 mm in diameter, the tumors were resected and cut into 1.5 mm-thick slices. RFP-expressing tumor stroma in the specimens was observed with the FV1000 confocal laser microscope (Olympus) [Uchugonova et al., 2011].

RESULTS AND DISCUSSION

ESTABLISHMENT OF A FLUORESCENT UPS PDOX MODEL

The PDOX tumor grew orthotopically in the right biceps femoris of an RFP expressing nude mouse. The PDOX tumor in the RFP-expressing nude mouse became brightly fluorescent (Fig. 1A). The resected PDOX tumor from the RFP transgenic nude mouse was also brightly fluorescent (Fig. 1B). The RFP expressing PDOX tumor was passaged orthotopically to the right biceps femoris of non-fluorescent nude mice to establish a non-invasively imageable PDOX model (Fig. 2). The bright RFP-expressing UPS PDOX was readily visible without opening the skin (Fig. 2A-left). The UPS PDOX tumor in the non-transgenic nude mouse was then exposed by a skin flap. Tumor RFP expression was very bright when imaged through the skin-flap (Fig. 2A-right). The resected RFP-expressing PDOX was also very bright (Fig. 2B).

CONFOCAL IMAGING OF THE STROMA IN THE UPS PDOX MODEL

FV1000 confocal laser microscope imaging showed RFP-expressing tumor stroma of PDOX tumors derived from RFP-expressing nude mice and after passage to a non-transgenic nude mouse. The PDOX model tumors had bright and diffusely distributed RFP expressing stroma (Fig. 3). Fibroblast-like cells and lymphocyte-like cells from the PDOX tumors were observed (Fig. 3).

UPS-ST5 PDOX tumors, previously grown in RFP transgenic nude mice for only one passage, had very bright fluorescence and after

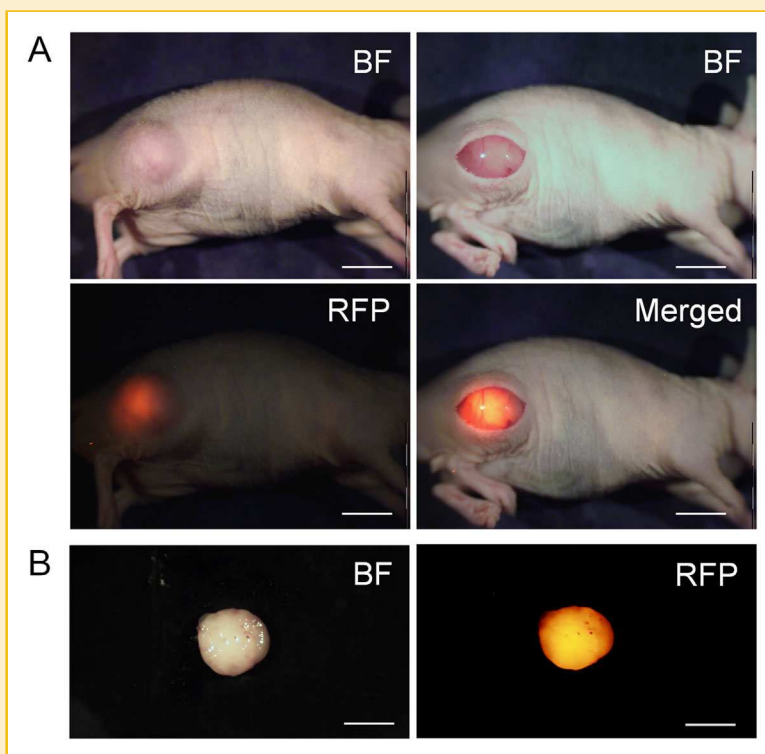


Fig. 2. (A) Non-invasive imaging of a RFP-expressing UPS PDOX tumor in a non-transgenic nude mouse, previously grown in an RFP-transgenic mouse (left). Imaging of the RFP-expressing UPS PDOX after a skin-flap was raised (right). (B) Images of the resected PDOX tumor derived from a non-transgenic nude mouse model. Tumor RFP expression was still strongly detectable. Imaging with the OV100. Scale bars: 10 mm (A and B). RFP, red fluorescent protein; BF, brightfield.

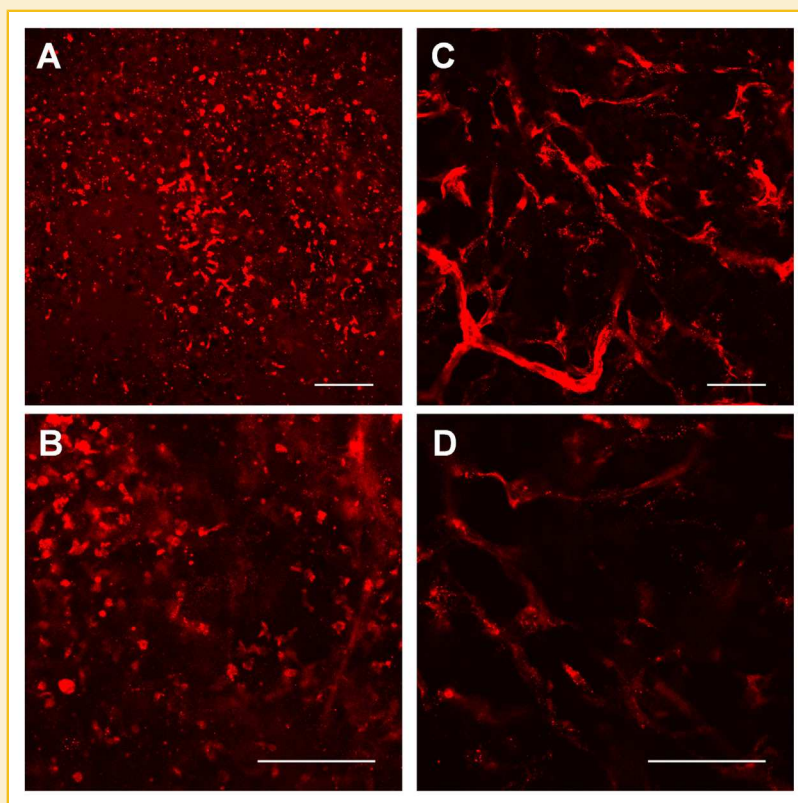


Fig. 3. Confocal fluorescence microscopy of individual stromal cells of the UPS PDOX. FV1000 confocal laser microscopy imaged RFP-expressing stroma of PDOX tumors grown in the RFP-expressing nude mouse (A and B) and after passage and growth in non-transgenic nude mice (C and D). Bright and diffusely-distributed RFP expressing stroma could be seen both in the UPS PDOX grown in the RFP transgenic nude mouse and after passage and growth in a non-transgenic nude mouse. Scale bars: 100 μ m.

passage to non-transgenic nude mice were non-invasively imageable due to maintenance of the RFP-expressing stroma within the PDOX. FV1000 confocal imaging demonstrated diffusely distributed bright RFP stromal cells in the PDOX tumor, both in RFP transgenic nude mice and after passage to non-transgenic nude mice. These results demonstrate a powerful method to make the PDOX UPS-STS model brightly fluorescent for non-invasive imaging, as well as for confocal microscopy of individual stromal cells associated with the tumor.

CONCLUSIONS

Implantation of human tumors in nude mice changed the paradigm of cancer research [Rygaard and Povlsen, 1969] and orthotopic implantation of tumor in nude mice [Sordat and Wang, 1984; Fidler, 1990; Fu et al., 1991; Hoffman, 2015] enabled patient-like models to be established for individualized patient therapy. In the present study, we established an imageable PDOX model of UPS-STS brightly labeled with RFP-expressing stroma in non-transgenic nude mice passaged from RFP transgenic nude mice. Only one passage in an RFP nude mouse was necessary to brightly and stably label the stroma of the UPS PDOX. The RFP-labeled UPS PDOX has the potential to rapidly screen for novel effective agents for individual patients, including stroma-targeting drugs, whereby the stromal cells are a visual target.

REFERENCES

- Astoul P, Wang X, Colt HG, Boutin C, Hoffman RM. 1996. A patient-like human malignant pleural mesothelioma nude-mouse model. *Oncol Rep* 3:483–487.
- Fidler IJ. 1990. Critical factors in the biology of human cancer metastasis: Twenty-eight G. H. A Clowes Memorial Award Lecture. *Cancer Res* 50:6130–6138.
- Fu X, Besterman JM, Monosov A, Hoffman RM. 1991. Models of human metastatic colon cancer in nude mice orthotopically constructed by using histologically intact patient specimens. *Proc Natl Acad Sci USA* 88:9345–9349.
- Fu X, Guadagni F, Hoffman RM. 1992. A metastatic nude-mouse model of human pancreatic cancer constructed orthotopically from histologically intact patient specimens. *Proc Natl Acad Sci USA* 89:5645–5649.
- Fu X, Hoffman RM. 1993. Human ovarian carcinoma metastatic models constructed in nude mice by orthotopic transplantation of histologically-intact patient specimens. *Anticancer Res* 13:283–286.
- Fu X, Le P, Hoffman RM. 1993. A metastatic-orthotopic transplant nude-mouse model of human patient breast cancer. *Anticancer Res* 13:901–904.
- Furukawa T, Kubota T, Watanabe M, Kitajima M, Fu X, Hoffman RM. 1993. Orthotopic transplantation of histologically intact clinical specimens of stomach cancer to nude mice: Correlation of metastatic sites in mouse and individual patient donors. *Int J Cancer* 53:608–612.
- Hiroshima Y, Zhao M, Maawy A, Zhang Y, Katz MH, Fleming BJ, Uehara F, Miwa S, Yano S, Momiyama M, Suetsugu A, Chishima T, Tanaka K, Bouvet M, Endo I, Hoffman RM. 2014a. Efficacy of *Salmonella typhimurium* A1-R

- versus chemotherapy on a pancreatic cancer patient-derived orthotopic xenograft (PDOX). *J Cell Biochem* 115:1254–1261.
- Hiroshima Y, Maawy A, Sato S, Murakami T, Uehara F, Miwa S, Yano S, Momiyama M, Chishima T, Tanaka K, Bouvet M, Endo I, Hoffman RM. 2014b. Hand-held high-resolution fluorescence imaging system for fluorescence-guided surgery of patient and cell-line pancreatic tumors growing orthotopically in nude mice. *J Surg Res* 187:510–517.
- Hiroshima Y, Maawy A, Zhang Y, Murakami T, Momiyama M, Mori R, Matsuyama R, Katz MH, Fleming JB, Chishima T, Tanaka K, Ichikawa Y, Endo I, Hoffman RM, Bouvet M. 2014c. Metastatic recurrence in a pancreatic cancer patient derived orthotopic xenograft (PDOX) nude mouse model is inhibited by neoadjuvant chemotherapy in combination with fluorescence-guided surgery with an anti-CA 19-9-conjugated fluorophore. *PLoS ONE* 9:e114310.
- Hiroshima Y, Zhang Y, Murakami T, Maawy AA, Miwa S, Yamamoto M, Yano S, Sato S, Momiyama M, Mori R, Matsuyama R, Chishima T, Tanaka K, Ichikawa Y, Bouvet M, Endo I, Zhao M, Hoffman RM. 2014d. Efficacy of tumor-targeting *Salmonella typhimurium* A1-R in combination with anti-angiogenesis therapy on a pancreatic cancer patient-derived orthotopic xenograft (PDOX) and cell line mouse models. *Oncotarget* 5:12346–12357.
- Hiroshima Y, Maawy AA, Katz MH, Fleming JB, Bouvet M, Endo I, Hoffman RM. 2015a. Selective efficacy of zoledronic acid on metastasis in a patient-derived orthotopic xenograft (PDOX) nude-mouse model of human pancreatic cancer. *J Surg Oncol* 111:311–315.
- Hiroshima Y, Maawy A, Zhan Y, Murakami T, Momiyama M, Mori R, Matsuyama R, Chishima T, Tanaka K, Ichikawa Y, Endo I, Hoffman RM, Bouvet M. 2015b. Fluorescence-guided surgery, but not bright-light surgery, prevents local recurrence in a pancreatic cancer patient-derived orthotopic xenograft (PDOX) model resistant to neoadjuvant chemotherapy (NAC). *Pancreatol* 15:295–301.
- Hiroshima Y, Zhao M, Zhang Y, Zhang N, Maawy A, Murakami T, Mii S, Uehara F, Yamamoto M, Miwa S, Yano S, Momiyama M, Mori R, Matsuyama R, Chishima T, Tanaka K, Ichikawa Y, Bouvet M, Endo I, Hoffman RM. 2015c. Tumor-targeting *Salmonella typhimurium* A1-R arrests a chemo-resistant patient soft-tissue sarcoma in nude mice. *PLoS ONE* 10:e0134324.
- Hiroshima Y, Zhang Y, Zhang N, Uehara F, Maawy A, Murakami T, Mii S, Yamamoto M, Miwa S, Yano S, Momiyama M, Mori R, Matsuyama R, Chishima T, Tanaka K, Ichikawa Y, Bouvet M, Endo I, Hoffman RM. 2015d. Patient-derived orthotopic xenograft (PDOX) nude mouse model of soft-tissue sarcoma more closely mimics the patient behavior in contrast to the subcutaneous ectopic model. *Anticancer Res* 35:697–701.
- Hoffman RM. 2015. Patient-derived orthotopic xenografts: Better mimic of metastasis than subcutaneous xenografts. *Nat Rev Cancer* 15:451–452.
- Kiyuna T, Murakami T, Tome Y, Kawaguchi K, Igarashi K, Zhang Y, Zhao M, Li Y, Bouvet M, Kanaya F, Singh A, Dry S, Eilber FC, Hoffman RM. 2016. High efficacy of tumor-targeting *Salmonella typhimurium* A1-R on a doxorubicin- and dactolisib-resistant follicular dendritic-cell sarcoma in a patient-derived orthotopic xenograft PDOX nude mouse model. *Oncotarget* 7:33046–33054.
- Murakami T, DeLong J, Eilber FC, Zhao M, Zhang Y, Zhang N, Singh A, Russell T, Deng S, Reynoso J, Quan C, Hiroshima Y, Matsuyama R, Chishima T, Tanaka K, Bouvet M, Chawla S, Endo I, Hoffman RM. 2016a. Tumor-targeting *Salmonella typhimurium* A1-R in combination with doxorubicin eradicate soft tissue sarcoma in a patient-derived orthotopic xenograft PDOX model. *Oncotarget* 7:12783–12790.
- Murakami T, Singh AS, Kiyuna T, Dry SM, Li Y, James A, W, Igarashi K, Kawaguchi K, DeLong JC, Zhang Y, Hiroshima Y, Russell T, Eckardt MA, Yanagawa J, Federman N, Matsuyama R, Chishima T, Tanaka K, Bouvet M, Endo I, Eilber FC, Hoffman RM. 2016. Effective molecular targeting of CDK4/6 and IGF-1 R in a rare FUS-ERG fusion CDKN2A-deletion doxorubicin-resistant Ewing's sarcoma in a patient-derived orthotopic xenograft (PDOX) nude-mouse model. *Oncotarget* Epub ahead of print.
- Rygaard J, Povlsen CO. 1969. Heterotransplantation of a human malignant tumour to "nude" mice. *Acta Path Microbiol Scand* 77:758–760.
- Sordat B, Wang WR. 1984. Human colorectal tumor xenografts in nude mice: Expression of malignancy. *Behring Inst Mitt* 74:291–300.
- Suetsugu A, Katz M, Fleming J, Truty M, Thomas R, Moriwaki H, Bouvet M, Saji S, Hoffman RM. 2012a. Multi-color palette of fluorescent proteins for imaging the tumor microenvironment of orthotopic tumorgraft mouse models of clinical pancreatic cancer specimens. *J Cell Biochem* 113:2290–2295.
- Suetsugu A, Katz M, Fleming J, Truty M, Thomas R, Saji S, Moriwaki H, Bouvet M, Hoffman RM. 2012b. Imageable fluorescent metastasis resulting in transgenic GFP mice orthotopically implanted with human patient primary pancreatic cancer specimens. *Anticancer Res* 32:1175–1180.
- Suetsugu A, Katz M, Fleming J, Truty M, Thomas R, Saji S, Moriwaki H, Bouvet M, Hoffman RM. 2012c. Non-invasive fluorescent-protein imaging of orthotopic pancreatic-cancer-patient tumorgraft progression in nude mice. *Anticancer Res* 32:3063–3067.
- Uchugonova A, Duong J, Zhang N, König K, Hoffman RM. 2011. The bulge area is the origin of nestin-expressing pluripotent stem cells of the hair follicle. *J Cell Biochem* 112:2046–2050.
- Wang X, Fu X, Hoffman RM. 1992. A new patient-like metastatic model of human lung cancer constructed orthotopically with intact tissue via thoracotomy in immunodeficient mice. *Int J Cancer* 51:992–995.
- Yamauchi K, Yang M, Jiang P, Xu M, Yamamoto N, Tsuchiya H, Tomita K, Moossa AR, Bouvet M, Hoffman RM. 2006. Development of real-time subcellular dynamic multicolor imaging of cancer cell trafficking in live mice with a variable-magnification whole-mouse imaging system. *Cancer Res* 66:4208–4214.
- Yang M, Reynoso J, Bouvet M, Hoffman RM. 2009. A transgenic red fluorescent protein-expressing nude mouse for color-coded imaging of the tumor microenvironment. *J Cell Biochem* 106:279–284.
- Yano S, Hiroshima Y, Maawy A, Kishimoto H, Suetsugu A, Miwa S, Toneri M, Yamamoto M, Katz MHG, Fleming JB, Urata Y, Tazawa H, Kagawa S, Bouvet M, Fujiwara T, Hoffman RM. 2015. Color-coding cancer and stromal cells with genetic reporters in a patient-derived orthotopic xenograft (PDOX) model of pancreatic cancer enhances fluorescence-guided surgery. *Cancer Gene Ther* 22:344–350.